Synthesis of C-2"β- and C-2"α-Fluoro Avermectin B_{1a}

Christophe Bliard, Francisca Cabrera Escribano, Gabor Lukacs,* Alain Olesker, and Pierre Sarda

Institut de Chimie des Substances Naturelles du CNRS, 91190 Gif-sur-Yvette, France

A synthesis of protected C-2 β - and C-2 α -fluoro oleandrosyl fluorides is presented; coupling of these carbohydrates with avermectin B_{1a} monosaccharide 5-*O*-t-butyldimethylsilyl derivative, followed by deprotection, furnished the title compounds whose biological activity was evaluated.

The avermectins constitute a group of eight closely related macrocyclic lactones with extremely potent antiparasitic activities.¹ Avermectin B_{1a} (1) is in development as an agricultural acaricide and insecticide. Structure-activity relationship studies¹ have shown that avermectin B_{1a} monosaccharide (2) has only about 25% and the aglycone alone about 5% of the biological activity of (1). These observations prompted us to undertake a programme aimed at further

improving the biological activity of avermectin B_{1a} by modifying the environment of its glycosidic linkage between the two oleandrose units. In order to make this glycosidic linkage more resistant to hydrolysis, we prepared C-2" β - and C-2" α fluoro avermectin B_{1a} , (**5a**) and (**5b**) respectively, and evaluated their biological activity.

Starting from L-rhamnose, benzyl 4-O-benzoyl-6-deoxy-3-O-methyl- α -L-mannopyranoside (6), a common protected



(4c)
$$R^1 = SiMe_2Bu^t$$
; $R^2 = Y$ with $R^3 = SiMe_2Bu^t$
(5c) $R^1 = H_1 R^2 + Y$ with $R^3 = \Gamma_1 R^4$

- (5a) $R^1 = H$; $R^2 = X$ with $R^3 = F$; $R^4 = R^5 = H$ (5b) $R^1 = H$; $R^2 = X$ with $R^3 = R^5 = H$; $R^4 = F$
- (5c) $R^1 = H; R^2 = Y$ with $R^3 = H$

synthetic intermediate of both C-2β- and C-2α-fluoro oleandrose, was prepared in an overall yield of 50%.² Hydrogenolysis of the benzyl group of (6) furnished (7) quantitatively. When the diol (7) was allowed to react at room temperature in tetrahydrofuran (THF) solution with 2.6 equiv. of diethylaminosulphur trifluoride (DAST)³ a mixture was produced consisting of the readily separable and useful (8) (62%) and (9) (34%) which upon acid hydrolysis was recycled to starting material (7). In the conversion of (7) into (8) the intermediate β -L-mannopyranosyl fluoride system (10) was expected to be highly favourable for such an S_N^2 process.⁴ However, the α -L-mannopyranosyl fluoride structure (9) remains unchanged in the presence of DAST even at higher temperatures. Base catalysed debenzoylation of (8) followed by t-butyldimethylsilylation gave quantitatively the appropriately protected C-2\beta-fluoro oleandrose derivative (11), activated at its anomeric centre and ready for the glycosylation step.5

The preparation of a protected C-2 α -fluoro oleandrose derivative was based on a recently discovered stereospecific 1,2-migration reaction in carbohydrate chemistry.^{6,7} DAST treatment of (6) in boiling benzene furnished an anomeric mixture (80%) of 4-O-benzoyl-2-O-benzyl-6-deoxy-3-Omethyl- α -L- (12) and - β -L-glucopyranosyl fluoride (13). Hydrogenolysis of the benzyl group of this mixture was followed by the preparation of the trifluoromethanesulphonates (14) and (15) which could be readily separated. In the presence of Bu₄N+F⁻ or CsF, as expected,⁴ fluorine introduction at C-2 on the β -L-glucopyranosyl fluoride system (15) proceeded better than on its α -L-counterpart (14) affording





respectively (17) (75%) and (16) (55%). Debenzoylation of (16) and (17) followed by t-butyldimethylsilylation gave quantitatively (18) and (19) respectively, ready for the glycosylation step.⁵

It is of interest to note the stability of C-1 fluoro-sugars in these and many other reactions which will be reported elsewhere.

In the presence of 1.5 equiv. of t-butyldimethylsilyl chloride, avermectin B_{1a} monosaccharide (2)¹ gave regiospecifically the 5-O-t-butyldimethylsilyl ether (3) (57%). Unreacted (2) (25%) could be also recovered. When a solution of (3) (1.15 mmol) in THF containing silver perchlorate, stannyl chloride, and molecular sieves was treated overnight in a nitrogen atmosphere, in the dark at +5 °C in three separate experiments with fluoro-sugars (11), (18), and (19) (1.24 mmol each), glycosylation products were obtained.⁵ Using (11), C-2"\beta-fluoro avermectin B_{1a} 4",5-bis-O-t-butyldimethylsilyl ether (4a) was produced ($\tilde{M^+}$ + H 1119) (18%). From (18) and (19), C-2" α -fluoro avermectin B_{1a} 4",5-bis-O-tbutyldimethylsilyl ether (4b) $(M^+ + H \ 1119)$ and the corresponding β -glycoside (4c) (M^+ + H 1119) were obtained after chromatographic separation. The glycosylation yield was about 20% and the ratio of (4b) to (4c) was 4:3 and 3:2 from respectively (18) and (19).

Structural proof for the glycosylation products was provided by their highly complex ¹³C n.m.r. spectra. One-bond, geminal, and vicinal ¹³C-¹⁹F coupling constants⁸ are in excellent agreement with the equatorial configuration of fluorine in (**4a**) and the axial orientation of fluorine in (**4b**) and (**4c**). The approximately identical chemical shift value of C-5" in the spectrum of (1)⁹ and (**4a**) (between δ 67.1 and 69.8) and (**4b**) (δ 69.8) fully supports the newly created glycosidic linkages as shown. In the spectrum of the β -glycoside (**4c**) the C-5" resonance is strongly deshielded (δ 73.5) as expected.

Deprotection of the C-4" and C-5 hydroxy groups of (4a), (4b), and (4c) with tetrabutylammonium fluoride, as described,² afforded (5a), (5b), and (5c) respectively, although in very moderate yields. This is presumably the result of the extreme sensitivity of the hydrogen atom at C-2 towards epimerisation under basic conditions.¹⁰ However, both silyl ether groups could be removed in excellent yields (85%) with the hydrogen fluoride-pyridine complex¹¹ giving the corresponding deprotected avermectins. ingly, the β -glycoside (**5c**) was found to be 500 times less active against helminths but equally effective against the two-spotted spider mite.

We are indebted to Merck, Sharp and Dohme for financial support and to Merck scientists Drs. M. H. Fisher and H. Mrozik for helpful discussions and to M. J. Wyvratt for suggesting the use of the hydrogen fluoride-pyridine complex in the last step of the synthesis. The mite and anthelmintic assay were done respectively by Drs. F. Preiser and D. Ostlind.

Received, 20th October 1986; Com. 1495

References

- 1 M. H. Fisher and H. Mrozik, in 'Macrolide Antibiotics,' ed. S. Omura, Academic Press, New York-London, 1984, p. 553.
- 2 K. C. Nicolaou, R. E. Dolle, D. P. Papahatjis, and J. L. Randall, J. Am. Chem. Soc., 1984, 106, 4189.
- 3 G. H. Posner and S. R. Haines, Tetrahedron Lett., 1985, 5.
- 4 M. Miljkovic, M. Gligorijevic, and D. Glisin, J. Org. Chem., 1974, 39, 3223.
- 5 T. Mukaiyama, Y. Murai, and S. Shoda, *Chem. Lett.*, 1981, 431. 6 S. Castillon, A. Dessinges, R. Faghih, G. Lukacs, A. Olesker, and
- T. That Thang, J. Org. Chem., 1985, **50**, 4913. 7 K. C. Nicolaou, T. Ladduwahetty, J. L. Randall, and A.
- Chucholowski, J. Am. Chem. Soc., 1986, 108, 2466. 8 V. Wray, J. Chem. Soc., Perkin Trans. 2, 1976, 1598.
- 9 G. Albers-Schönberg, B. H. Arison, J. C. Chabala, A. W. Douglas, P. Eskola, M. H. Fisher, A. Lusi, H. Mrozik, J. L. Smith, and R. L. Tolman, J. Am. Chem. Soc., 1981, 103, 4216.
- 10 J. V. Pivnichny, J.-S. K. Shim, and L. A. Zimmerman, J. Pharm. Sci., 1983, 72, 1447.
- 11 K. C. Nicolaou, S. P. Seitz, M. R. Pavia, and N. A. Petasis, J. Org. Chem., 1979, 44, 4011; K. C. Nicolaou, S. P. Seitz, and M. R. Pavia, J. Am. Chem. Soc., 1981, 103, 1222.